Lignans from the Bark of Magnolia kobus

by Hak-Ju Lee^a), Seon-Mi Seo^a), Oh-Kyu Lee^a), Hyun-Jin Jo^a), Ha-Young Kang^a), Don-Ha Choi^a), Ki-Hyon Paik^b), and Merajuddin Khan*^a)

- a) Division of Wood Chemistry and Microbiology, Korea Forest Research Institute, Seoul 130-712, Korea (phone: +82-2-961-2747; fax: +82-2-961-2747; e-mail: mdk_cimap@yahoo.com)
- b) Department of Forest Resources and Environmental Science, Korea University, Seoul 136-701, Korea

The Et₂O-soluble fraction from the bark of *Magnolia kobus* led to the isolation of two new lignans, (+)- $(7\alpha,7'\alpha,8\alpha,8'\alpha)$ -3'-4,4'-5,5'-pentamethoxy-7,9':7'-9-diepoxylignan-3-ol (1) and (+)- $(7\alpha,7'\alpha,8\alpha,8'\alpha)$ -4,5-dimethoxy-3'-4'-(methylenedioxy)-7,9':7'-9-diepoxylignan-3-ol (2), along with five known lignans 3-7. Their structures were established on the basis of various spectroscopic analyses including 1D- (1 H, 13 C, and DEPT) and 2D-NMR (COSY, NOESY, HMQC, and HMBC) and by comparison of their spectral data with those of related compounds.

Introduction. – Magnolia kobus DC belongs to the Magnoliaceae family. It is a medium-sized deciduous tree native to Japan also found in China and Korea [1]. It is a valuable decorative plant in Japan and is known under the local name Kobusi. Young buds of *M. kobus* are important ingredients in the Chinese medicine 'Shin-I' which is used as a sedative or an analgesic. In Japan, 'Shin-I' is taken internally for the treatment of headaches or colds [2]. Earlier chemical studies on *M. kobus* revealed it to be a source of bioactive terpenes and lignans [3–6]. Lignans have evoked a great deal of interest due to their widespread occurrence in nature [7–9] and use in traditional medicines [10][11]. Furofuran lignan, one of the major subclasses of the lignan family, exhibit a wide variety of biological activities including antitumor, antimitomic, antiviral [12], antioxidant, antihypertensive [13][14], and antidiabetic activity [15], and inhibition of the platelet-activating factor (PAF) [16].

In the present study, two new furofuran lignans, (+)- $(7\alpha,7'\alpha,8\alpha,8'\alpha)$ -3',4,4',5,5'-pentamethoxy-7,9':7',9-diepoxylignan-3-ol (1) and (+)- $(7\alpha,7'\alpha,8\alpha,8'\alpha)$ -4,5-dimethoxy-3',4'-(methylenedioxy)-7,9':7',9-diepoxylignan-3-ol (2), along with five known compounds, (+)-sesamin (3), (+)-yangambin (4), (+)-kobusin (5), (+)-eudesmin (6), and (+)-magnolin (7), were isolated from the bark of M. kobus (Fig. 1). Their structures were established on the basis of various spectroscopic analyses including 1D- (1 H, 13 C, and DEPT) and 2D-NMR (COSY, NOESY, HMQC, and HMBC) and by comparison of their spectral data with those of related compounds.

Results and Discussion. – Extensive chromatographic separation and purification of the Et_2O -soluble fraction from the EtOH extract of *M. kobus* bark led to the isolation of the two new furofuran lignans **1** and **2**, along with the five known lignans **3**–**7**.

Compound 1 was obtained as white crystals and displayed a molecular-ion peak at m/z 432 (M^+) in the EI-MS. Its molecular formula could be determined as $C_{23}H_{28}O_8$ by

Fig. 1. Compounds 1-7, isolated from the bark of M. kobus

its HR-EI-MS peak at m/z 432.1459 (M^+). The ¹H- and ¹³C-NMR signals of **1** (*Table*) were assigned by the interpretation of the DEPT, COSY, NOESY (*Fig.* 2), HMQC, and HMBC data (*Fig.* 2). On the basis of these spectroscopic evidences and of comparison with lignans of the furofuran type [17–21] and of the (+)-sesamin type [18][22][23], **1** was characterized as (+)-(7α , $7'\alpha$, 8α , $8'\alpha$)-3',4,4',5,5'-pentamethoxy-7,9':7',9-diepoxylignan-3-ol, *i.e.*, with (7S,7'S,8R,8'R) configuration (*Fig.* 1).

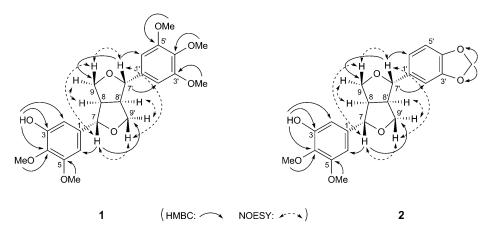


Fig. 2. Key NOE and HMBC of compounds 1 and 2

The 13 C-NMR spectrum of **1** showed the signals for 23 C-atoms which were distinguished by DEPT experiments as five Me, two CH₂, and eight CH groups, and eight quaternary C-atoms. The 1 H-NMR spectrum of **1** showed signals for two CH groups at δ (H) 3.07 – 3.15 (m, 2 H), two benzylic OCH moieties at δ (H) 4.73 (d, J = 4.5 Hz, 1 H) and 4.75 (d, J = 4.5 Hz, 1 H), two oxygenated CH₂ groups at δ (H) 4.27 – 4.35 (m, 2 H) and 3.91 – 3.94 (m, 2 H), and two 1,3,4,5-tetrasubstituted benzene rings at δ (H) 6.53 (d, J = 2.0 Hz, 1 H) and 6.59 (s, 3 H), which were assigned to a lignan of the furofuran type by comparison with data reported previously [17][18]. This observation was further confirmed by the corresponding 13 C-NMR signals at δ (C) 54.08 and 54.31 (2 CH), 85.60 and 85.99 (2 benzylic OCH), at 71.73 and 72.15 (2 oxygenated CH₂), 134.84, 136.71, 137.32, 137.45, 149.31, 152.50, and 153.39 (8 aromatic C), and 101.67, 102.81, 105.46 (4 aromatic CH), as established by DEPT and 1 H, 13 C one-bond (HMQC) experiments. The 1 H-NMR spectrum showed the signals for a OH group at δ (H) 5.86 (br. s) and five MeO groups at δ (H) 3.84, 3.88, and 3.90 (3s), all being substituents at benzene rings. The 1 H- and 13 C-NMR data of **1** was

Table. ${}^{1}H$ - and ${}^{13}C$ -NMR Data (CDCl₃; 500 and 125 MHz, resp.) of Compounds 1-2. δ in ppm,

Position	1		2	
	$\delta(C)$	$\delta(H)$	$\delta(C)$	$\delta(\mathrm{H})$
C(1)	137.32	_	137.41	_
H-C(2)	105.46	6.59(s)	105.48	6.57 (d, J = 1.5)
C(3)	149.31	_	149.31	-
C(4)	134.84	_	134.83	-
C(5)	152.50	_	152.50	-
H-C(6)	101.67	6.53 (d, J = 2.0)	101.66	6.50 (d, J = 1.5)
H-C(7)	85.99	4.75 (d, J = 4.5)	85.77	4.72 (d, J = 3.0)
H-C(8)	54.08	3.07-3.15 (m)	54.26	3.06-3.11 (m)
$CH_2(9)$	72.15	4.27-4.35 (m), 3.91-3.94 (m)	71.74	4.25-4.30 (m), 3.90-3.92 (m)
C(1')	136.71	_	135.00	_
H-C(2')	102.81	6.59(s)	106.48	6.86 (s)
C(3')	153.39	_	147.95	_
C(4')	137.45	_	147.10	_
C(5') or $H-C(5')$	153.39	_	108.16	6.80 (d, J = 8.0)
H-C(6')	102.81	6.59(s)	119.35	6.82 (d, J = 8.0)
H-C(7')	85.60	4.73 (d, J = 4.5)	85.66	4.71 (d, J = 3.0)
H-C(8')	54.31	3.07-3.15 (m)	54.16	3.06-3.11 (m)
$CH_2(9')$	71.73	4.27-4.35 (m), 3.91-3.94 (m)	71.91	4.25-4.30 (m), 3.90-3.92 (m)
OCH ₂ O	_	_	101.04	5.96 (s)
OH-C(3)	_	5.86 (br. s)	_	5.84 (s)
MeO-C(4)	60.91	3.90 (s)	60.92	3.89(s)
MeO-C(5)	55.89	3.88(s)	55.89	3.88 (s)
MeO-C(3')	56.15	3.88 (s)	_	_
MeO-C(4')	60.80	3.84 (s)	_	_
MeO-C(5')	56.15	3.88 (s)	-	-

very similar to that of (+)-yangambin (4) which was also isolated and characterized in the present study. The presence of an OH group at C(3) of 1 instead of an MeO group in 4 was confirmed by the 2D-NMR data, i.e., by the HMBCs $\delta(H)$ 5.86 (br. s, OH)/C(3), C(2) and C(4) (Fig. 2). The equivalence of the $\delta(H)$ of H-C(7) and H-C(7), H-C(8) and H-C(8), and H-C(9) and H-C(9) required a symmetrical relative configuration for the aryl substituents [18] [19] which could be either axial, axial or equatorial, equatorial. The equatorial, equatorial position of the aryl substituents was assigned on the basis of the following features of the ¹H- and ¹³C-NMR spectra [18-21]: i) The chemical shifts of the fusion sites C(8) and C(8') of 1 were δ (C) 54.08 and 54.31, respectively, while for furofuran lignans with diaxial aryl substituents, C(8) and C(8') appear more upfield (δ (C) 49.0 – 49.5). ii) The chemical shifts of $CH_2(9)$ and $CH_2(9')$ of **1** were $\delta(H)$ 4.27 – 4.35 $(m, 2H, H_a)$, and at 3.91 – 3.94 $(m, 2H, H_\beta)$, while these protons appear more upfield at $\delta(H)$ 3.65-4.0 $(m, 2H, H_a)$ and 3.30-3.65 $(m, 2H, H_\beta)$ if the aryl substituents are axial. iii) Small coupling constants were observed for H-C(7) and H-C(7') (J=4.5 Hz) of 1. This observation was further confirmed by the NOEs $H_{ax}-C(7')/H_{ax}-C(9)$ and $H_{ax}-C(9')$ and $H_{ax}-C(7)/H_{ax}-C(9)$ and $H_{ax}-C(9)$ (Fig. 2) confirming that the aryl substituents are diequatorial. According to Freudenberg and Sidhu [22], all sesamin-type lignans with a positive $[\alpha]_D$ have the same absolute configuration (R) at the fusion sites C(8) and C(8'). Hence, **1** with an $[a]_D$ of +42.6 should have (8R,8'R)-configuration. Moreover, the aryl substituents of 1 being diequatorial and $[\alpha]$ being positive, the (7S,7'S)-configuration of 1 is confirmed. Indeed all (+)-(8R,8'R)-sesamin type-lignans with diequatorial aryl substitution belong to the same series with the absolute configuration (S) at the benzylic C(7) and C(7') atoms [18][23].

Compound **2** was obtained as white crystals and exhibited a molecular-ion peak at m/z 386 (M^+) in the EI-MS. Its molecular formula could be established as $C_{21}H_{22}O_7$ from the HR-EI-MS peak at m/z 386.1374 (M^+). Most of the 1H - and ^{13}C -NMR signals (Table) of **2** were similar to those of **1**. On the basis of dextrorotatory nature and relative configuration, the absolute configuration of **2** must be the same as that of **1** [18–20][22][23]. Therefore, the structure of **2** was established as (+)-(7α , $7'\alpha$, 8α , $8'\alpha$)-4,5-dimethoxy-3',4'-(methylenedioxy)-7,9':7',9-diepoxylignan-3-ol with (7S,7'S,8R,8'R) configuration (Fig. 1).

The ¹H-NMR signal of **2** at δ (H) 5.96 (s, 2 H) and its corresponding ¹³C-NMR signal at δ (C) 101.04, established from an HMQC experiment, suggested a OCH₂O group in the molecule. The major changes observed in the ¹H- and ¹³C-NMR spectra of **2** as compared to **1** are: i) appearance of an extra CH signal at δ (H) 6.80 (d, J = 7.5 Hz, H–C(5')); ii) more downfield shifted signals for H–C(6') and H–C(2' (δ (H) 6.82 (d, J = 8.5 Hz) and 6.86 (s, resp.) and for C(6') and C(2') (δ (C) 119.35 and 106.48, resp.); iii0 downfield and upfield shifted quaternary C-atom signals (δ (C) 147.10 (C(4')) and 147.95 (C(3')). The OCH₂O group was located at C(3') and C(4') based on the HMBC δ (H) 5.96 (s, OCH₂O)/C(3') and C(4') (Fig. 2). The [a]_D of **2** was positive, and the relative configuration of the two aryl substituents were established as equatorial, equatorial on the basis of spectral features similar to those of **1**.

The known compounds 3-7 (Fig. 1) were identified by comparison of their spectral data with literature values as follows: (+)-sesamin (3) [5][24], (+)-yangambin (4) [23][25], (+)-kobusin (5) [5][19], (+)-eudesmin (6) [5][19], and (+)-magnolin (7) [17][26].

The authors thank the Korea Basic Science Institute in Seoul for performing the NMR experiments.

Experimental Part

General. TLC: precoated silica gel 60 F_{254} (SiO₂, 0.2 mm; Merck) plates; developed with solvent system A, toluene/HCOOEt/HCOOH 5:4:1 and 7:2:1 (v/v/v) and visualized under UV light at 254 and 365 nm, resp. Column chromatography (CC): SiO₂ 60 (40–100 μm; Kanto Chemical Co.); Adventec-SF-1600 automated fraction collector. M.p.: Electrothermal-IA-9200 melting point apparatus (Electrothermal Engg. Ltd., U. K.). Optical rotations: Jasco-P-1020 polarimeter. UV Spectra: Hewlett-Packard-8452A diode-array spectrophotometer; λ_{max} (log ε) in nm. IR Spectra: Nexus-FT-IR spectrophotometer; $\tilde{\nu}$ in cm⁻¹. ¹H- and ¹³C-NMR Spectra: Varian-Unity-Inova-500 spectrophotometer; at 500 (¹H) and 125 MHz (¹³C); in CDCl₃; δ in ppm rel. to Me₄Si as internal standard, J in Hz. EI and HR-EI-MS: Jeol-JMS-SX102A spectrophotometer; in m/z (rel. %).

Plant Material. The bark of M. kobus was collected from the experimental forest of the Southern Forest Research Center, Kyungnam, Korea, in June 2001, and identified by Dr. Y. H. Kwon (Korea National Arboretum, Pocheon, Korea). A voucher specimen was deposited with the Korea Forest Research Institute, Seoul, Korea.

Extraction and Isolation. Shade air-dried and powdered barks (12.0 kg) of *M. kobus* were extracted three times with 95% EtOH at r.t. for 72 h each. The combined EtOH extracts were concentrated under vacuum at 40° until EtOH was completely removed. The concentrated EtOH extract was dissolved in dist. H₂O and successively partitioned with petroleum ether, Et₂O, and AcOEt. The Et₂O-soluble fraction (85.0 g) was subjected to CC (*Sephadex LH-20*, MeOH/EtOH 3:7): *Fractions 1* and 2. *Fr. 2* was subjected to CC (SiO₂, benzene/AcOEt 8:1): *Frs. 2.1–2.8. Fr. 2.2* gave pure **3** (500 mg). *Fr. 2.3* (2.52 g) was subjected to CC(SiO₂, CHCl₃/MeOH 20:1: *Fr. 2.3.1–2.3.5. Fr. 2.3.4* (758.9 mg) was subjected to two repeated CC (SiO₂, CHCl₃/MeOH 100:1): pure **2** (24 mg). *Frs. 2.4* and 2.5 were subjected separately to CC (SiO₂, hexane/AcOEt 2:1); *Sephadex*, MeOH: pure **5** (1.25 g) and **6** (120 mg), resp. *Fr. 2.6* (20.1 g)

was subjected to CC (SiO₂, hexane/AcOEt 2:1): Frs. 2.6.1 – 2.6.3. Fr. 2.6.3 (15.3 g) was again subjected to CC (Sephadex LH-20, EtOH): Frs. 2.6.3.1 – 2.6.3.3. Fr. 2.6.3.1 (1.0 g) gave, on CC (SiO₂, CHCl₃/MeOH 100:1), Frs. 2.6.3.1.1 – 2.6.3.1.6. Frs. 2.6.3.1.4 (240 mg), 2.6.3.1.5 (250 mg), and 2.6.3.1.6 (200 mg) were separately subjected to CC (SiO₂, CHCl₃/MeOH 80:1): pure 7 (30 mg), 4 (25 mg), and 1 (16 mg), resp.

 $(+)\cdot (7\alpha,7'\alpha,8\alpha,8'\alpha)\cdot 3',4,4',5,5'-Pentamethoxy\cdot 7,9':7',9-diepoxylignan-3-ol \qquad (=2,3-Dimethoxy\cdot 5-\{(IS,3aR,4S,6aR)\cdot tetrahydro\cdot 4\cdot (3,4,5-trimethoxyphenyl)\cdot IH,3H-furo[3,4-c]furan-1-yl]phenol; 1): White crystals. M.p. <math>116-118^\circ$. $[\alpha]_2^{D.4}=+42.6$ (c=0.10, CHCl $_3$). UV (CHCl $_3$): 235 (3.9), 273 (3.6). IR (KBr): 3417, 2927, 2359, 1599, 1515, 1465, 1412, 1264, 1235, 1141, 1056, 1025. 1 H- and 1 C-NMR: *Table*. EI-MS: 432 (M^+ , 100), 207, 195, 181, 167, 161. HR-EI-MS: 432.1459 (M^+ , $C_{23}H_{28}O_8^+$; calc. 432.1784).

(+)- $(7\alpha,7'\alpha,8\alpha,8'\alpha)$ -4,5-Dimethoxy-3',4'-(methylenedioxy)-7,9':7',9-diepoxylignan-3-ol (=5-[(1S,3aR,4S,6aR)-4-(1,3-Benzodioxol-5-yl)tetrahydro-1H,3H-furo[3,4-c]furan-1-yl]-2,3-dimethoxyphenol; **2**): White crystals. M.p. 129 – 131°. [α] $_{0}^{20.4}$ = +56.2 (c = 0.14, CHCl $_{3}$). UV (CHCl $_{3}$): 248 (4.1), 276 (3.7). IR (KBr): 3414, 2924, 2356, 1594, 1508, 1463, 1440, 1245, 1236, 1138, 1045, 1026. 1 H- and 13 C-NMR: *Table*. EI-MS: 386 (M⁺, 100), 195, 181, 169, 161, 149, 135, 122, 83. HR-EI-MS: 386.1374 (M⁺, C_{21} H $_{22}$ O $_{7}^{+}$; calc. 386.1366).

- (+)-Sesamin (3): White crystals. $[\alpha]_D^{20.4} = +62.0$ (c = 0.58, CH₂Cl₂) ([24]: $[\alpha]_D = +63.2$). EI-MS: 354 (M^+). UV, IR, ¹H- and ¹³C-NMR: in agreement with [5][24].
- (+)-Yangambin (4): White solid. $[\alpha]_D^{20.4} = +43.2$ (c = 0.42, CHCl₃) ([25]: $[\alpha]_D = +45.1$). EI-MS: 446 (M^+). UV, IR, ¹H- and ¹³C-NMR: in agreement with [23][25].
- (+)-Kobusin (5): Colorless oil. $[\alpha]_D^{20.4} = +56.8$ (c = 0.54, CH_2Cl_2) ([19]: $[\alpha]_D = +58.0$). EI-MS: 370 (M^+). UV, IR, ¹H- and ¹³C-NMR: in agreement with [5][19].
- (+)-Eudesmin (6): Amorphous powder. $[\alpha]_0^{20.4} = +59.9$ (c = 0.59, acetone) ([19]: $[\alpha]_D = +61.0$). EI-MS: 386 (M^+). UV, IR, ¹H- and ¹³C-NMR: in agreement with [5][19].
- (+)-Magnolin (7): White solid. $[\alpha]_D^{20.4} = +55.1$ (c = 0.07, MeOH) ([26]: $[\alpha]_D = +55.7$). EI-MS: 386 (M^+). UV, IR, ¹H- and ¹³C-NMR: in agreement with [17][26].

REFERENCES

- [1] T. B. Lee, 'Illustrated Flora of Korea', Hwang Mun Sa, Seoul, Korea, 1982, p. 374.
- [2] H. Matsutani, T. Shiba, Phytochemistry 1975, 14, 1132.
- [3] Y. Fijuta, M. Kikuchi, S. Fijuta, Yakugaku Zasshi 1975, 95, 235.
- [4] H. Hirose, M. Satoh, A. Hagitani, Nippon Kagaku Kaishi 1968, 89, 889.
- [5] T. Iida, M. Nakano, K. Ito, Phytochemistry 1982, 21, 673.
- [6] J. Li, M. Tanaka, K. Kurasawa, T. Ikeda, T. Nohara, J. Nat. Med. 2007, 61, 222.
- [7] J. Lee, D. Lee, D. S. Jang, J.-W. Nam, J.-P. Kim, K. H. Park, M. S. Yang, E.-K. Seo, Chem. Pharm. Bull. 2007, 55, 137.
- [8] R. S. Ward, Chem. Soc. Rev. 1982, 11, 75.
- [9] R. S. Ward, Nat. Prod. Rep. 1999, 16, 75.
- [10] E. Okuyama, K. Suzumura, M. Yamazaki, Chem. Pharm. Bull. 1995, 43, 2200.
- [11] M. Takasaki, T. Konoshima, I. Yasuda, T. Hamano, H. Tokuda, Biol. Pharm. Bull. 1997, 20, 776.
- [12] Y. M. Chiung, H. Hayashi, H. Matsumoto, T. Otani, K. Yoshida, M. Y. Huang, R. X. Chen, J. R. Liu, M. Nakayama, J. Antibiot. 1994, 47, 487.
- [13] R. C. Brown, C. J. Bataille, G. Bruton, J. D. Hinks, N. A. Swain, J. Org. Chem. 2001, 66, 6719.
- [14] W. D. MacRae, G. H. N. Towers, Phytochemistry 1984, 23, 1207.
- [15] C.-H. Hou, S.-J. Lin, J.-T. Cheng, F.-L. Hsu, J. Nat. Prod. 2003, 66, 625.
- [16] S. Iwakami, J. B. Wu, Y. Ebizuka, U. Sankawa, Chem. Pharm. Bull. 1992, 40, 1196.
- [17] M. Miyazawa, H. Kasahara, H. Kameoka, Phytochemistry 1993, 32, 1421.
- [18] A. Pelter, R. S. Ward, E. V. Rao, K. V. Sastry, Tetrahedron 1976, 32, 2783.
- [19] J. Latip, T. G. Hartley, P. G. Waterman, Phytochemistry 1999, 51, 107.
- [20] D. C. Ayres, J. D. Loike, 'Lignans: Chemical, Biological and Clinical Properties (Chemistry and Pharmacology of Natural Products)', Cambridge University Press, Cambridge, 1990, p. 209.

- [21] B.-S. Min, M.-K. Na, S.-R. Oh, K.-S. Ahn, G.-S. Jeong, G. Li, S.-K. Lee, H. Joung, H.-K. Lee, J. Nat. Prod. 2004, 67, 1980.
- [22] K. Freudenberg, S. Sidhu, Chem. Ber. 1961, 94, 851.
- [23] H. Greger, O. Hofer, *Tetrahedron* **1980**, *36*, 3551.
- [24] L. Jayasinghe, B. M. M. Kumarihamy, K. H. R. N. Jayarathna, N. W. M. G. Udishani, B. M. R. Bandara, N. Hara, Y. Fujimoto, *Phytochemistry* **2003**, *62*, 637.
- [25] W. D. MacRae, G. H. N. Towers, *Phytochemistry* **1985**, 24, 561.
- [26] H. Kakisawa, T. Kusumi, H. Y. Hsu, Y. P. Chen, Bull. Chem. Soc. Jpn. 1970, 43, 3631.

Received June 5, 2008